Thin-layer chromatography of imino-acids

Imino-acids in plants are of three main types, based on the 4-, 5-, and 6-membered ring systems of azetidine, pyrrolidine and piperidine, respectively. Azetidine-2carboxylic acid is widely distributed in the Liliaceae and occurs in a few species of Agavaceae, but is otherwise a rare plant component¹. Of the pyrrolidine compounds, 4-hydroxyproline is a constituent of a few plant proteins, and proline is of widespread occurrence. Several piperidine compounds have been isolated from plants, including piperidine-2-carboxylic acid^{2,3}, 5-hydroxypiperidine-2-carboxylic acid⁴, 4-hydroxypiperidine-2-carboxylic acid⁵ and 1,2,3,6-tetrahydropyridine³. In several of the papers describing the characterisation of these piperidine compounds, evidence has been obtained by comparing the chromatographic characteristics of the unknown compound with synthesised reference compounds²⁻⁶. The chromatographic methods described have been with paper, but the solvent systems used have given only small differences in R_F values between the compounds, and hence the time of development required for effective separation of the compounds was lengthy. In recent work on Salix fragilis L. leaf galls, a new imino-acid was encountered, probably based on piperidine. For the study of this compound, thin-layer chromatographic methods were developed which gave discrete and rapid separations of the different imino-acids.

Experimental and results

Air dried silica gel G (Merck) layers, 250μ thick, on glass plates 20×20 cm were used. The test compounds, in aqueous solution, were applied near one corner of the plate, the spot was dried with a hot air blower and developed using the super-saturated method of STAHL⁷. Two-way chromatograms were prepared using solvent systems that have been used successfully for the separation of amino-acids⁹. For the first direction, development was with chloroform-methanol-17 % ammonia (2:2:1, v/v), and for the second direction the solvent used was either (a) propanol-water (64:36, w/w), (b) *n*-butanol-acetic acid-water (60:20:20, w/w), or (c) phenol-water (4:1, w/w). Development in each direction was for 15 cm from the point of application of the spot. The spots were detected with an 0.1 % w/v solution of ninhydrin in acetone and with an 0.2 % w/v solution of isatin in *n*-butanol containing 4 % acetic acid. The colours were developed by heating the plates at 100° for 10 min (Table I). The compounds examined were piperidine-2-, piperidine-3-, and piperidine-4-carboxylic acids;

TABLE I

IMINO-ACID COLOURS PRODUCED WITH ISATIN AND NINHYDRIN

Compound	Isatin	Ninhydrin
Piperidine-2-carboxylic acid	blue-green	mauve to blue
Piperidine-3-carboxylic acid	blue-green	mauve to blue
Piperidine-4-carboxylic acid	blue-green	mauve to blue
4-Hydroxypiperidine-2-carboxylic acid	weak green	yellow changing to blue
5-Hydroxypiperidine-2-carboxylic acid	blue-green	mauve to blue
Compound from Salix fragilis galls	blue-green	mauve to blue
4-Hydroxyproline	blue-green	yellow changing to pink
Proline	green-blue	yellow

4-hydroxy- and 5-hydroxypiperidine-2-carboxylic acids; 4-hydroxyproline, proline and an imino-acid from *Salix fragilis* leaf galls, probably based on piperidine. Using the two-way chromatographic systems described, all the imino-acids tested were easily separated and clearly identified (Fig. 1).

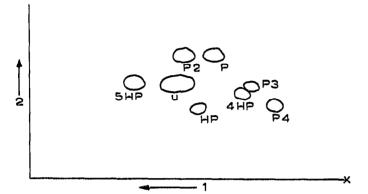


Fig. 1. Thin-layer chromatogram of imino-acids. 5HP = 5-hydroxypiperidine-2-carboxylic acid; 4HP = 4-hydroxypiperidine-2-carboxylic acid; P2 = piperidine-2-carboxylic acid; P3 = piperidine-3-carboxylic acid; P4 = piperidine-4-carboxylic acid; P = proline; HP = 4-hydroxyproline; u = compound from S. *fragilis* galls. Solvent systems; (A) chloroform-methanol-17% ammonia (2:2:1, v/v) and (B) phenol-water (4:1, w/w).

Acknowledgements

We wish to thank Prof. A. VIRTANEN for the gift of samples of 4-hydroxy- and 5-hydroxypiperidine-2-carboxylic acid and Prof. CLARK-LEWIS for a sample of 4-hydroxypiperidine-2-carboxylic acid.

School of Pharmacy, Portsmouth College of Technology, Portsmouth (Great Britain) GERALD BLUNDEN STEPHEN B. CHALLEN

۰.

I L. FOWDEN, Abhandl. Deut. Akad. Wiss. Berlin, Kl. Chem. Geol. Biol., No. 4 (1963) 17.

- 2 R. I. MORRISON, Biochem. J., 53 (1953) 474.
- 3 N. GROBBELAAR, J. K. POLLARD AND F. C. STEWARD, Nature, 175 (1955) 703.
- 4 A. I. VIRTANEN AND S. KARI, Acta Chem. Scand., 8 (1954) 1290.
- 5 A. I. VIRTANEN AND S. KARI, Acta Chem. Scand., 9 (1955) 170.
- 6 L. FOWDEN, Biochem. J., 70 (1958) 629.
- 7 E. STAHL, Arch. Pharm., 292 (1959) 411.

8 E. STAHL, Thin Layer Chromatography, Springer-Verlag, Berlin, 1965, p. 399.

Received February 25th, 1966

J. Chromatog., 24 (1966) 224-225